

## Expression and Purification of pancreatic lipase-like protein

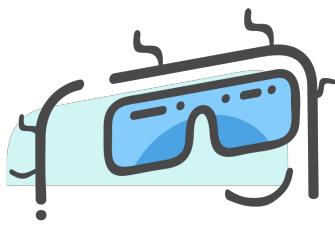
### Gene and Expression Vector

#### 1.1 Gene coding for the target protein

The cDNA coding for the target protein was chemically-synthesized with optimization for expression in *E.coli* and described as below.

##### > *16\_pancreatic\_lipase-like\_protein-1445bp*

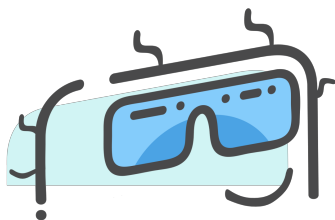
atgggtcaccatcaccatcaccatgtgtcgactcagaagtcaatcaagaagctaagccagagggtca  
agccagaagtcaagcctgagactcacatcaatttaaagggtgtccgatggatcttcagagatcttcttcaagat  
caaaaagaccactccttaagaaggctgatggaagcggtcgctaaaagacagggttaaggaaatggactcc  
ttaagattctgtacgacggtattagaattcaagctgatcagaccctgaagatttgacatggaggataac  
gatattattgaggctcacagagaacagattggtggccaaggatccATGGCCGAGGTGTGCTACGG  
TGACCTGGGCTGCTTCACCAACATTGCGCCGTTTTGGGGCCTGAACCGTCCGCTGA  
GCAGCCTGCCGGACCCGCCGGATGTGGTTGGTACCAAGTTCTACCTGTATACCCGTG  
CGAACCCGACGATGGCGCTGCGTGAGCGTCTGGTGGCGGATAGCATCGCGACCCTG  
AGCGCGAGCCACTTCAGCAGCAGCAAGCCGACCCGTATGGTTATCCACGGTTTTAC  
CGGTAGCGCGGAACATGCGTGGGTGCAGACCATTGTTGACGAGCTGCTGCTGAAAG  
AAGATATCAACGTGATTACCGTTGACTGGGCGGATGGCGCGAGCATTGGTGGCAGC  
TACGGTCAAGCGACCGCGAACAGCCGTGTGGTTGGTGCGGAAGTGGCGAAGATTG  
TTAACTATATGAGCGCGCAAACCGGTGCGAACGCGCGTAACTCCACCTGATTGGTC



ACAGCCTGGGTTGCCATGTGGCGGGTTATGCGGGTGACATCCTGGGTAACGTTGGC  
CGTATTACCGCGCTGGATGCGAGCGAACCGTATTTTGACGGTATGGATGCGATCGTG  
AAACTGGACCCGACCGATGCGCTGTTCGTGGACGTTATCCACAGCGATGGTAGCCCG  
TTTATTGGTACCCTGGGTATGGGTACCAGCCTGCCGACCGGTCATGTGGACTTCTACC  
CGAACGGTGGCGAGTATCAGCCGGGTTGCCACGATAACTTCGTTAGCAGCGTGTT  
AGCACCGGTTTTGGTCTGCTGACCGAGGGTTATGATGGTGCGGAAGCGGCGGCGGC  
GTGCAGCCACCTGCGTGCGATCGATTATTTACCGAGAGCATTAAACAGCGAATGCCC  
GTTTACCGCGTACCCGTGCGAAAGCTATGACAAGTTCAAAGATGGTTTTTGCATGAG  
CTGCGCGGGTAGCACCTGCAGCCAAATGGGTTACCGTGCGCGTGACCACTATGGTG  
CGCGTGGCAAACGTACCTGACCACCACCGATGGTACCCAGCCGGGTGGCAAGTTC  
TGCGCGTACCACTATAAAGTGCAAGTTACCAGCAGCACCCGTATGGAGGAAACCGC  
GGGTCGTATCTACGTTACCTTTTCAGGGTCCGAGCGGCATTACCGACACGATGGAAGT  
GACCGACGGCGATGTTATGGATCTGAAGCCGGGTGTGACCTATCAAAAAGTGGTTAG  
CAGCGCGAGCAACATCGGTGACGTGAGCCGTGTTGGCCTGCGTTTCGAACGTAGCA  
GCAGCATTTGGGATTTCTTTGCGGCGCAGTACTATACCGTGGGCAAGGTTACCATCA  
CCGCGGGCGAGACCCAAACCAGCTACAAATTTTGCGGCAGCGACACCCCGGTGAA  
CAACGGTGAAACCCTGCAGCTGACCCAAGTTGGTGGCACCTGCtgaCTCGAG

## 1.2 Expression vector

The cDNA sequence described was cloned in pATX-sumo expression vector.



> *16\_pancreatic\_lipase-like\_protein (581AAs,62.98kDa, pl:5.05)*

MGHHHHHHMSDSEVNQEAKPEVKPEVKPETHINLKVSDGSSEIFFKIKKTTPLRR  
LMEAFAKRQKGKEMDSLRFYDGIRIQADQTPEDLDMEDNDIIEAHREQIGGQGSMAE  
VCYGDLGCFTNIAPFWGLNRPLSSLPDPPDVVGTKFYLYTRANPTMALRERLVADSIAT  
LSASHFSSSKPTRMVIHGFTGSAEHAWVQTIVDELLKEDINVITVDWADGASIGGSYG  
QATANSRVVGAEVAKIVNYMSAQTGANARNFHLIGHSLGCHVAGYAGDILGNVGRIT  
ALDASEPYFDGMDAIVKLDPTDALFVDVIHSDGSPFIGTLGMGTSLPTGHVDFYPNGG  
EYQPGCHDNFVSSVVSTGFGLLTEGYDGAEEAAACSHLRAIDYFTESINSECPFTAYPC  
ESYDKFKDGFCSMAGSTCSQMgyrardHYGARGKLYLTtTDGTQPGGKFCAYHYK  
VQVTSSTRMEETAGRIYVTFQGPSGITDTMEVTDGDVMDLKPGVTYQKLVSASNIG  
DVSRVGLRFRERSSSIWDFFAAQYYTVGKVTITAGETQTSYKFCGSDTPVNNGETLQLTQ  
VGGTC

Features: SUMO-tag:[1:109]

## Subcloning

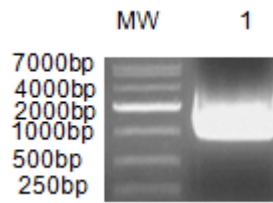
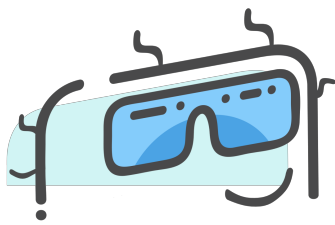
### 2.1 Primer

9408p-4-F gaacagattggtggccaaggatccATGGCCGAGGTGTGCTACGGTGAC

9408p-4-R tcagtgggtggtggtggtgctcgagtcaGCAGGTGCCACCAACTGGGTCAGCTG

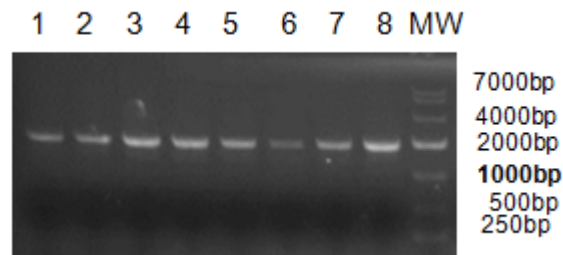
PCR reaction system	50ul
2x PrimeSTAR MAX	25ul
Forward Primer: 10uM	2ul
Reverse Primer:10uM	2ul
Template:	0.5ul
ddH2O	to 50ul

PCR profile	
Denaturation	95℃,10s
Annealing	55℃,5s
Extension	72℃, 15s
Cycles	30 times
Storage	16℃,hold



**Figure 1. PCR analysis for target gene fragment. Goldview staining.**

Target gene was obtained. Subcloning test was done, the gene was collected with pATX-sumo vector and the positive clones were identified with PCR. The results were showed as below.



**Figure 2. PCR analysis for positive clones. Goldview staining.**

## 2.2 Conclusion of the test

Sequencing results showed that positive clone was obtained.

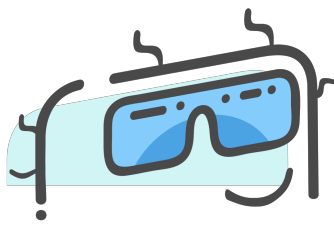
## ● Expression Tests

### 3.1 Aim

-Determination of optimal conditions for protein expression by evaluation of:

\*Induction strategy

\*Temperature and time for induction



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\**E.coli* strain used for protein production

-Identification of best extraction condition: native or denatured

### 3.2 Short protocol description

-Culture:

\*Bacteria starter obtained by incubation at 37°C

\*IPTG induction with different temperatures and incubation times (see details in §3.3).

-Protein samples preparation (for each expression condition tested):

\*Bacteria harvested by centrifugation

\*Lysis with native buffer

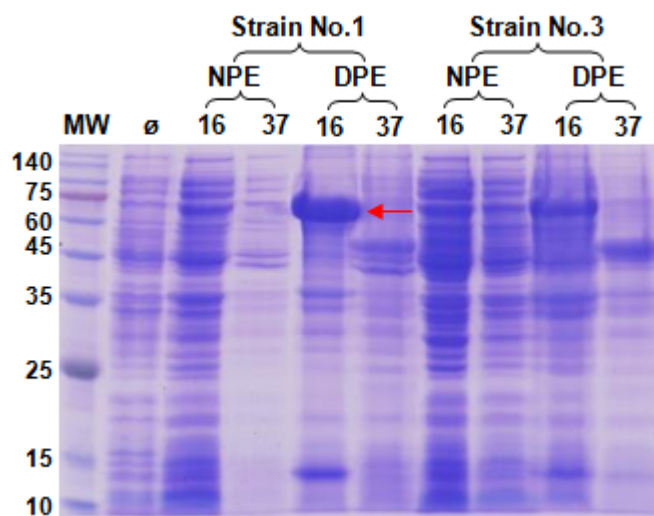
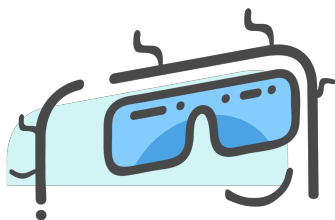
\*Clarification (centrifugation C1): The supernatant is the native protein extract (NPE)

\*Solubilization of the pellet from C1 with a denaturing buffer (Urea 8M)

\*Clarification (centrifugation): The supernatant is the denatured protein extract (DPE)

\*PAGE analysis of NPE and DPE.

### 3.3 Results of the expression tests



**Figure 3. Expression tests of the target protein.** Reducing SDS-PAGE.

Analysis of NPE and DPE prepared as described in §3.2.

**MW.** Molecular weight marker. **Ø.** Non-induced bacteria culture (negative control).

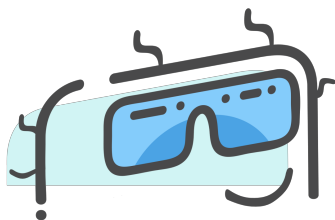
**16 and 37.** Incubation temperature (°C) during induction with IPTG.

Induction with IPTG 1mM during 16h at 16°C, or during 4h for other temperatures.

### 3.4 Conclusion of the expression tests

**Note:** *E.coli* strains tested are intended to overcome one or several expression issues such as (but not limited to) expression level, solubility or toxicity.

Target proteins had expressed, but most were in denatured condition.



Optimal expression condition was described as below.

	Strain No.	Temperature	Induction Time
<b>16 pancreatic lipase-like protein (DPE)</b>	No.1	37°C	4h

**Table1. Optimal denatured expression conditions of target protein.**

## 1L scale-up purification tests

### 4.1 Aim

1L scale up purification tests in denatured conditions for target protein.

Starting material: DPE prepared after production in optimal denatured expression conditions described in the Table 1.

### 4.2 Short protocol description

-Lysis buffer: PBS, pH7.5, 10% Glycerol

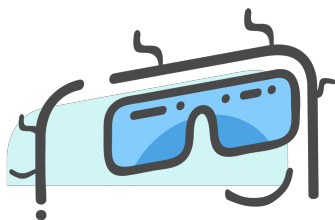
-Washing buffer: PBS, pH7.5., 1% Triton X 100, 5 mM EDTA

-Denaturing buffer: PBS, pH7.5, 2% Sarcosyl

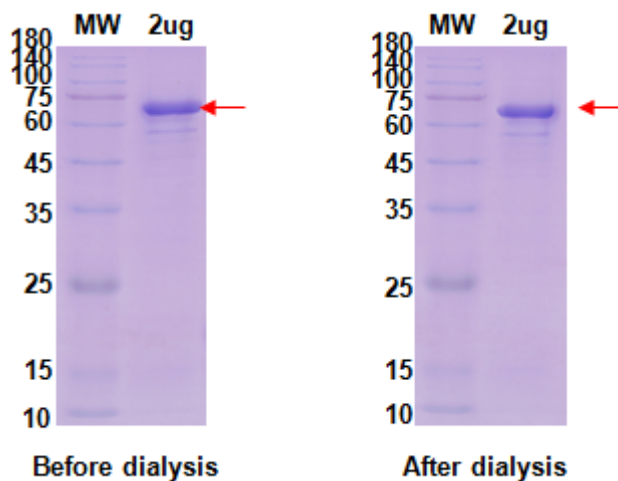
-Dialysis buffer: PBS, pH7.5, 0.02% Sarcosyl

-Analysis by SDS-PAGE of fractions of interest

-Final sample QC: qualitative by SDS-PAGE, quantitative by Bradford method.



#### 4.3 Results of the purification tests



**Figure 4. Final sample QC of denatured condition.** *Coomassie blue staining.*

Reducing-PAGE analysis. 2 $\mu$ g of sample loaded.

#### 4.4 Conclusion of the purification tests

Target proteins could be obtained from DPE.