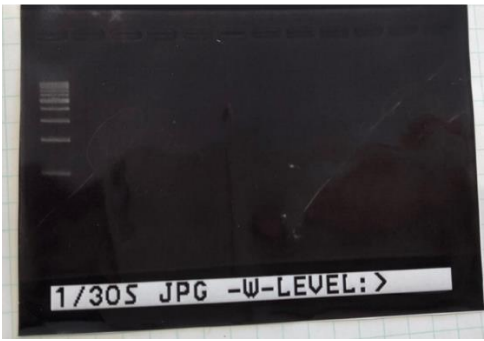


0923

1. Electrophoresis

1 2 3 4



1 One STEP Ladder Marker 500

2 pSB1C3

3 pSB1C3

4 pSB1C3

2. Pre-culture

3. PCR

Reagent

one sample

materials	volume( $\mu$ L)
KOD One Pol	25
template DNA	1.0
forward primer	1.5
reverse primer	1.5
D.W	20
total	49

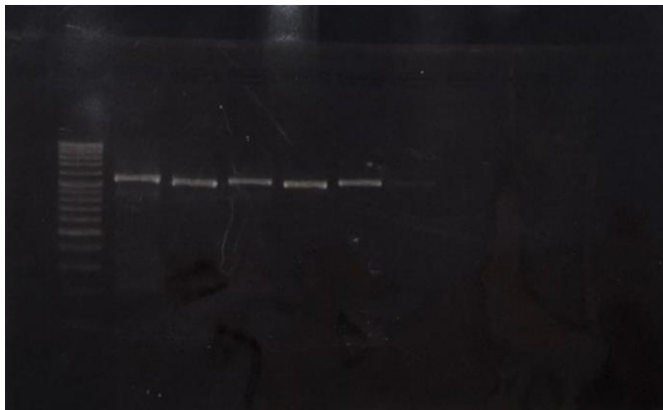
sample	forward primer	reverse primer	template
1	I	tsu	10-fold dilution
2	VII	X	10-fold dilution
3	I	tsu	100-fold dilution
4	VII	X	100-fold dilution
5	I	tsu	dH <sub>2</sub> O
6	VII	tsu	dH <sub>2</sub> O

#### Cycling condition

reaction		temp.(°C)	time
cycle 1 (× 1)	step 1	98	2:00
cycle 2 (× 30)	step 1	98	0:10
	step 2	68	1:00
cycle 3 (× 1)	step 1	4	∞

#### 4. Electrophoresis

1 2 3 4 5 6 7



- 1 One STEP Ladder Maker 100
- 2 Homologous sequence addition
- 3 recA
- 4 Homologous sequence addition
- 5 recA
- 6 Negative control
- 7 Negative control

0924

## 1. Colony PCR

### Reagent

materials	volume (μL)
D.W.	22
KOD One	25
forward primer	1.5
reverse primer	1.5
total	50

Sample	forward primer	reverse primer	colony
1	h	g	vector RFP 1
2	h	g	vector RFP 2
3	VII	X	insert 9/24 Colony1
4	VII	X	insert 9/24 Colony2
5	VII	X	insert 9/24 Colony3
6	h	g	vector 9/24 Colony4
7	h	g	vector 9/24 Colony5
8	h	g	vector 9/23 Colony6

### Cycling condition

reaction		temp.(°C)	time
cycle 1 (X1)	step 1	98	2:00
cycle 2 (X30)	step 1	98	0:10
	step 2	68	2:20
cycle 3 (X1)	step 1	4	∞

## 2. Electrophoresis

1 2 3 4 5 6 7 8 9 10 11 12



1 One STEP Ladder Marker 500

2 vector(RFP)

3 vector(RFP)

4 vector

5 vector

6 vector

7 One STEP Ladder Marker 100

8 recA

9 recA

10 recA

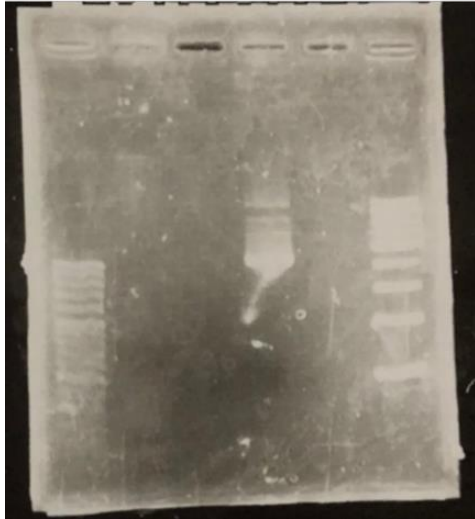
11 recA

12 recA

0925

1. Plasmid extraction

1      2              3              4



1 One STEP Ladder Maker 100

2 recA

3 vector(RFP)

4 One STEP Ladder 500

2. Making culture medium LB agar plate

0926

1. Electrophoresis

1      2              3              4



1 One STEP Ladder Maker 100

2 One STEP Ladder Maker 500

3 recA

4 pSB1C3

## 2. Transformation

## 3. Colony PCR

### Reagent

materials	volume (μL)
D.W.	22
KOD One	25
forward primer	1.5
reverse primer	1.5
total	50

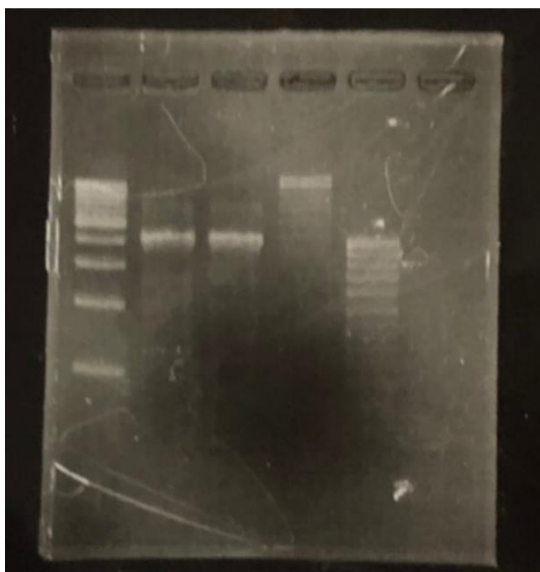
Sample	forward primer	reverse primer	colony
1	h	g	RFP
2	h	g	RFP
3	VII	X	RFP

### Cycling condition

reaction		temp.(°C)	time
cycle 1 (X1)	step 1	94	2:00
cycle 2 (X30)	step 1	98	0:10
	step 2	58	0:05
	step 3	68	0:12
cycle 3 (X1)	step 1	4	∞

#### 4. Electrophoresis

1 2 3 4 5



1 One STEP Ladder Maker 500

2 pSB1C3

3 pSB1C3

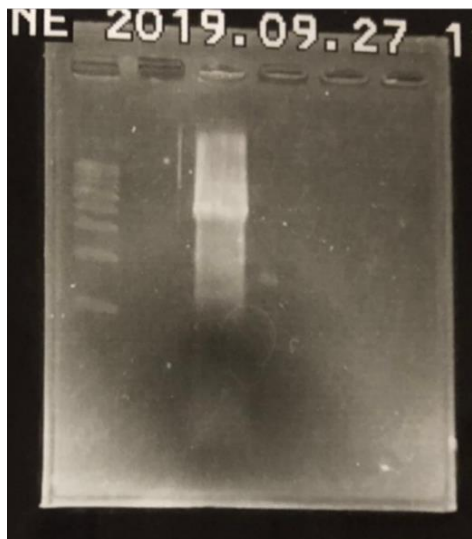
4 recA

5 One STEP Ladder Maker 100

0927

#### 1. Electrophoresis

1 2



1 One STEP Ladder Maker 500

2 pSB1C3

3. Colony PCR

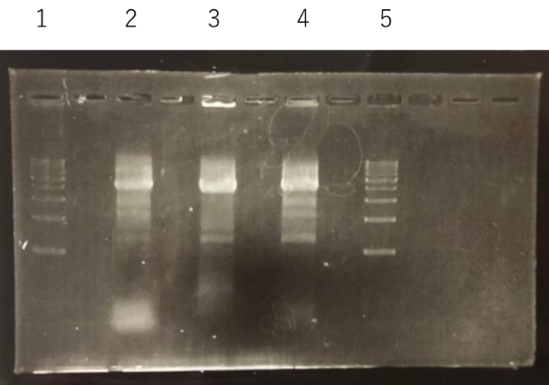
Reagent

materials	volume (μL)
D.W.	22
KOD One	25
forward primer	1.5
reverse primer	1.5
total	50

Cycling condition

reaction		temp.(°C)	time
cycle 1 (×1)	step 1	94	2:00
cycle 2 (×30)	step 1	98	0:10
	step 2	68	0:11
cycle 3 (×1)	step 1	4	∞

Electrophoresis



1 One STEP Ladder Maker 500

2 pSB1C3

3 pSB1C3(RFP)

4 pSB1C3(RFP)

5 One STEP Ladder Maker 500



#### 4. PCR for homologous sequence addition

##### Reagent

materials	volume ( $\mu\text{L}$ )
D.W.	21
KOD One	25
forward primer	1.5
reverse primer	1.5
template	1
total	50

sample	forward primer	reverse primer	template
1	recA for pSB1C3+recA	recA for pSB1C3+recA	10-fold dilution
2	recA for pSB1C4+recA	recA for pSB1C4+recA	10-fold dilution
3	recA for pSB1C5+recA	recA for pSB1C5+recA	100-fold dilution
4	recA for pSB1C6+recA	recA for pSB1C6+recA	100-fold dilution
5	recA for pSB1C7+recA	recA for pSB1C7+recA	dH <sub>2</sub> O
6	recA for pSB1C8+recA	recA for pSB1C8+recA	dH <sub>2</sub> O

##### Cycling condition

reaction		temp.(°C)	time
cycle 1 ( $\times 1$ )	step 1	98	2:00
cycle 2 ( $\times 30$ )	step 1	98	0:10
	step 2	68	0:16
cycle 3 ( $\times 1$ )	step 1	4	$\infty$