



Modelling – Lipase activity test

● Aim

To test whether the enzymes expressed by our group are active, we designed a series of enzyme activity experiments. Lipase activity experiments are an important part of this.

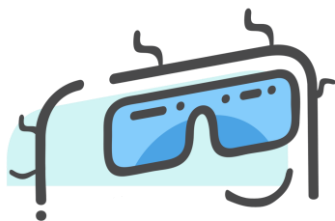
● Materials

1. Lipase Activity Assay Kit, Catalog Number MAK046 (Sigma-Aldrich);
2. Pancreatic lipase-like protein (serial number 16);
3. Transparent 96-well plate

● Procedure

Based on the instructions in the Lipase Activity Assay Kit, we designed the relevant experiments as follows:

1. Set Glycerol Standards for Colorimetric Detection: Dilute 10 μL of the 100 mM Glycerol Standard with 990 μL of the Lipase Assay Buffer to prepare a 1 mM standard solution. Add 0, 2, 4, 6, 8, and 10 μL of the 1 mM standard solution into a 96 well plate, generating 0 (blank), 2, 4, 6, 8, and 10 nmole/well standards. Add Lipase Assay Buffer to each well to bring the volume to 50 μL .
2. Sample Preparation: For the rigor and scientific nature of the



experiment, we divided our enzymes into three concentrations for each experiment, and each concentration was subjected to three parallel replicate experiments. The concentration gradients we set are: $1\times$ (0.017mg/mL), $10\times$ (0.17mg/mL), $100\times$ (1.7mg/mL);

3. Assay Reaction

(1) Set up the Reaction Mixes according to the scheme in Table 1. 100 μ L of the Reaction Mix is required for each reaction (well). Note: Glycerol in the samples will generate a background signal. To remove the effect of glycerol background, a Sample Blank may be set up for each sample by omitting the Lipase Substrate.

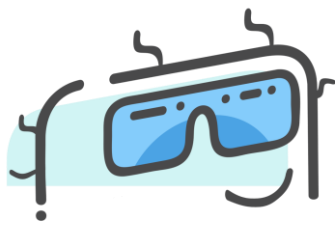
| Reagent | Standards and Samples | Sample Blank |
|----------------------|-----------------------|--------------|
| Lipase Assay Buffer | 93 mL | 96 mL |
| Peroxidase Substrate | 2 mL | 2 mL |
| Enzyme Mix | 2 mL | 2 mL |
| Lipase Substrate | 3 mL | — |

Table 1. Reaction Mixes

(2) Add 100 μ L of the appropriate Reaction Mix to each of the wells. Mix well by pipetting.

(3) Incubate the plate at 37°C. After 2–3 minutes (T_{initial}), measure the absorbance at 570 nm [$(A_{570})_{\text{initial}}$]. Note: It is essential [$(A_{570})_{\text{initial}}$] is in the linear range of the standard curve.

(4) Continue to incubate the plate at 37°C measuring the absorbance



$[(A_{570})_{\text{final}}]$ after 120 minutes. Protect the plate from light during the incubation.

(5) Correct for the background by subtracting the final measurement

$[(A_{570})_{\text{final}}]$ obtained for the 0 (blank) glycerol standard from the final measurement $[(A_{570})_{\text{final}}]$ of the standards and samples. Background values can be significant and must be subtracted from all readings. Plot the glycerol standard curve.

