Nanopore Sequencing

Preparing sample:

- Mix 9 µl of each sample with 1 µl of index.
- Incubate in thermocycler with the following settings:
  - 1 minute at 30°C
  - 1 minute at 80°C
- Pipet all samples in a reaction tube.
- Add 40 µl AM pure XP (beads) to buffer.
- Put in magnet separator and remove the remaining suspension.
- Carefully wash with alcohol two times, removing the remaining suspension quickly each time.
- Centrifuge briefly and remove the rest of the remaining alcohol.
- Dissolve in 10 µl water and mix by snapping the reaction tube gently. Do not let the beads dry out.
- Put in magnet separator.
- Add 1 µl rapid adapter (RAP) in a new reaction tube.
- Add 10 µl of the beads and water mix to the RAP-mix.
- Incubate for 10 minutes

Preparing the flow cell:

- Add 30 µl FLT in a tube of Flush Buffer (FB)
- Open the priming port and try to remove air from the flow cell by pipetting.
- Add 750 µl Flush Buffer inside.

Loading the samples:

- Mix 35 µl sequencing buffer with 25 µl loading beads in a reaction tube.
- Load 150 µl flush buffer in the priming port.
- Load the samples at the spot on.

From: iGEM Bielefeld-CeBiTec