

## IMPACT™ Kit: Protein Purification from NEB

### 1. Bacteria Cultivation:

- Inoculate an overnight culture with 25 mL LB and the required antibiotic in a 250 mL culture flask. Incubate at 37°C.
- Start a new culture with 250 mL LB and the required antibiotic in a 1 L culture flask from the overnight culture with start-OD<sub>600</sub> = 0.1
- Cultivate at 37°C until the OD-value reaches 0.5 – 0.8.
- Initiate expression by adding 0.4 M IPTG and let the bacterial growth proceed at 37°C for 30 minutes.
- Expression of the protein takes place at 17°C overnight. Use shaker in cooling chamber.
- Harvest the cells in a centrifuge with cooling functions for 20 minutes on 4,500 rpm at 4°C in a 250 mL centrifuge container.
- Decant the supernatant and resuspend in 35 mL lysis buffer.
- Transfer the resuspension in a 50 mL tube with screw cap.

### 2. Cell Disruption:

- The cells are to be disrupted in lysis buffer in a French Press® with the pressure of 1,212 Psi.
- Repeat the disruption 2 – 3 times.
- Centrifuge the cells for 1 hour on 4,500 rpm and 4°C. transfer the supernatant into a new 50 mL tube.

### 3. Column preparation:

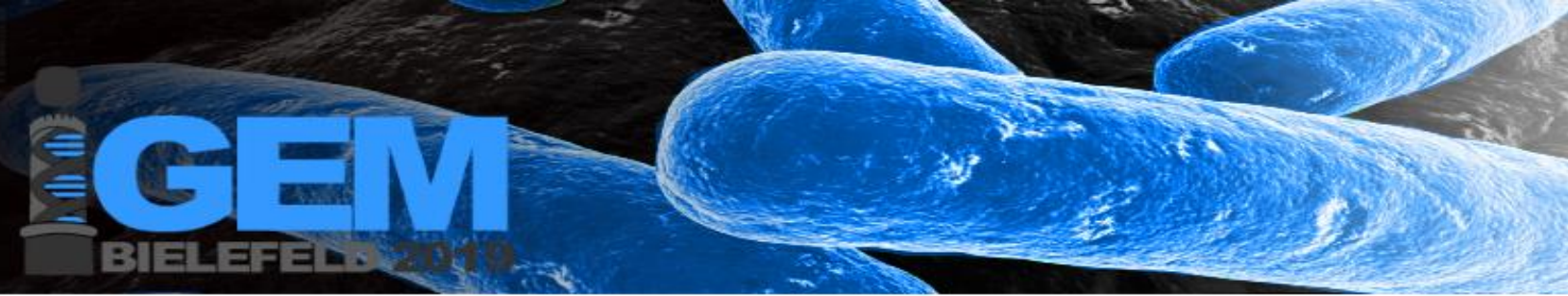
- Provide a rubber hose and a clamp to the bottom part of the column.
- Fill the column with 10 mL chitin-beads.
- Wait until the beads drop to the bottom and add the filter to the column material.
- Remove ethanol through the filter.
- Equilibrate the column with 10x column buffer.

### 4. Loading the column:

- Load the column with the supernatant from the cell disruption (max. 0.5 – 1.0 mL/min)
- Wash the column with 10x column-wash-buffer

### 5. Cleavage Initiation:

- Provide fresh cleavage buffer under no-light conditions (cover with tinfoil)
- Run 15 mL of the cleavage buffer through the column.
- Let sit at room temperature in the dark for 20 – 24 hours.



## 6. Elution:

- After the cleavage, elute with 1x column buffer.
- Concentrate the protein solution with the suitable protein filter.
- Wash the protein 3 times each with 10 mL column buffer.
- The end volume of the eluat: 250 – 500  $\mu$ L
- Create a 1:1 mixture of the eluat with 86% glycerin and store at  $-20^{\circ}\text{C}$ .

### Lysis Buffer:

- Tris 25 mM
- NaCl 500 mM
- EDTA 1 mM
- Triton X-100 0.15%
- PMSF 20  $\mu$ M
- TCEP 1mM

Adjust pH to 8.5

### Column-Wash-Buffer:

- Tris 25 mM
- NaCl 1500 mM
- EDTA 1mM

Adjust pH to 8.5

### Column Buffer:

- Tris 25 mM
- NaCl 500 mM
- EDTA 1 mM

Adjust pH to 8.5

### Cleavage Buffer:

- Tris 25 mM
- NaCl 500 mM
- EDTA 1 mM
- DTT 50 mM

Adjust pH to 8.5

From: iGEM Bielefeld-CeBiTec