



## COLOR CODING KEY

**Characterization**

**Construct**

**Data Collection/Analysis**

**Improvement of Biobrick**

**Plant-Care**

**Plasmid**

**Plasmid/Construct Design**

**Preparatory Work**

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### **Sunday June 23rd**

- Streaked 14 plates with bacteria that contained PreLim plasmid (Amp resistance)
  - Incubated plates at 37 C for overnight
  - Poured Agar Plates, added Agar to regular LB broth, mixed (note it was a little clumpy and took a while to dissolve)
  - Sterilized bench, water bath, cool, added Kanamycin
  - Did not cool and harden (probably because we did not autoclave after adding the agar)
  - Put into hot bath to cool to 50-60 degrees
  - Added Kanamycin
  - Poured Plates and let them solidify
  - Streaked 1 plate with bacteria that contained main project plasmid ( Kan resistance)
  - Incubated at 37C for overnight
  - Incubated empty plate as negative control
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### **Monday June 24th**

- Made 250 ml of agar liquid
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### **Tuesday June 25th**

- Streaked 8 plated with PreLim plasmid, and 2 control plates

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**Wednesday June 26th**

- Created a detailed Outline for Characterization with Constitutive Promoter (Bronze - BBa\_K541503)
  - GFP (experiment 1)
  - Bradford Assay (experiment 2)
- Liquid inoculation of PreLim plasmids
- Plated KAN plates (main project)

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**Thursday June 27th**

- Potted plants that were grown on 6/7
- Pruned and removed the buds of the plants
- Minipreped PreLim plasmid
  - Low concentration, but good 260/280 numbers
  - Changed protocol to fix this issue (water instead of elution buffer, 6 ul instead of 30 ul, no dilution)
- Liquid inoculation of KAN plates
- Made 1.2% agarose gel

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**Friday June 28th**

- Minipreped DNA for PreLim backbone
  - Low 260/230 numbers, due to not doing optional carbohydrate and phenol filtering step
- Problems with samples floating out of the gel during gel electrophoresis

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**Saturday June 29th**

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