

## Transformation of DinIII GFP into *O. marina* with Use of Lonza 4D-Nucleofector X Electroporation Unit System in 16 -well Nucleocuvette Strips

\* University of Connecticut (UConn) fed the *O. marina* *D. tertiolecta* 3 days prior to transformation. **Deviation:** The *O. marina* culture was fed 2 days prior. To add, UConn used a flashlight to concentrate the *O. marina* within the light. This seems weird as from my experience, the *O. marina* tend to swim away when under the microscope light.

1. For each transformation well, 16.4  $\mu\text{L}$  of solution SG, 3.6  $\mu\text{L}$  of Supplemental 1 solution, and 2  $\mu\text{L}$  of linear DNA product were used as a transformation solution.
2. *O. marina* cells were collected in a 50 mL falcon tube. The cells were centrifuged a 2500 g for 3 minutes.
3. Removed remaining medium supernatant except for 2-3 mL.
4. Transferred cells into 1.5 mL Eppendorf tubes and centrifuged at 900 g for 2 minutes. Discarded all remaining supernatant.
5. Cells were resuspended in transformation solution (reference step 1.) and 22  $\mu\text{L}$  were added into each well.
  - a. Added  $2.5 \times 10^5$  of *O. marina* cells into each well. **Deviation:** They calculated the *O. marina* cell density using a Sedgwick-Rafter counting chamber. However, we do not have an estimated cell density.
6. After an initial optimization test, the following electroporation settings were used for further experiments: DS-137, DS-130, DS-138, DS-134, DS-150, ED-150, DS-120, and no pulse controls (NPC).
7. Immediately after electroporation, 80  $\mu\text{L}$  of the same medium that *O. marina* was cultured in along with an AKS antibiotic cocktail (100  $\mu\text{g}/\mu\text{L}$  ampicillin, 50  $\mu\text{g}/\mu\text{L}$  kanamycin, and 50  $\mu\text{g}/\mu\text{L}$  streptomycin) was added into each well. **Deviation:** UConn grew their *O. marina* cultures in autoclaved, filtered seawater. We grew ours in F2 medium + autoclaved, filtered seawater.
8. All of the volume was gently transferred into 24-well plates where each well already contained 1.4 mL of the same F2 SW+AKS medium.
9. The transformed cells were left to recover for 3 days.
10. For the DinIII-gfp transformations, cells were examined microscopically under blue light for gfp expression.
11. New antibiotic solution was added every 3 weeks but the concentration was dropped down to 200  $\mu\text{g}/\mu\text{L}$  to allow for greater cell growth, and *D. tertiolecta* in 225  $\mu\text{g}/\mu\text{L}$  rifampin medium was supplied whenever they were no longer detected in the medium.
  - a. The *D. tertiolecta* in 225  $\mu\text{g}/\mu\text{L}$  rifampin medium was added to UConn's DinIII-arrO cells, so we added the *D. tertiolecta* within the F2 medium.